



Susan Crowley
CEM-1428

(702) 651-2234
Fax (405) 302-4607
susan.crowley@tronox.com

February 11, 2010

Mr. Brian Rakvica
Nevada Division of Environmental Protection
2030 E. Flamingo Road, # 230
Las Vegas, Nevada 89119-0818

**Subject: Technical Memo - Protocol: Bioaccessibility Method for Dioxin/Furans in Soil
Tronox LLC, Henderson Nevada: NDEP Facility ID # 000539**

Dear Mr. Rakvica:

Please find attached the subject technical memo.

I hereby certify that I am responsible for the services described in this document and for the preparation of this document. The services described in this document have been provided in a manner consistent with the current standards of the profession and, to the best of my knowledge, comply with all applicable federal, state and local statutes, regulations and ordinances.

Please contact me at (702) 592-7727 if you have any comments or questions concerning this correspondence. Thank you.

Sincerely,

Susan M. Crowley, CEM 1428, exp 3-8-11

Federal Express

CC: Shannon Harbour, NDEP
Jim Najima, NDEP
Paul Black, Neptune
Paul Hackenberry, Hackenberry
Teri Copeland
Dave Gratson, Neptune
Joanne Otani-Fehling, Neptune
Tom Reed, Tronox
Mike Logan, Tronox
Matt Paque, Tronox
Keith Bailey, Environmental Answers
Deni Chambers, Northgate
Renee Kalmes, Exponent

Tronox. Adding value beyond the product.

560 West Lake Mead Parkway, Henderson, Nevada 89015 • P.O. Box 55, Henderson, Nevada 89009

From: Deni Chambers, Northgate
Renee Kalmes, Exponent

DATE: February 11, 2010

To: Brian Rakvica, NDEP

RE: Protocol: Bioaccessibility Method for Dioxin/Furans in Soil

As requested By NDEP in their February 9, 2010 memorandum, the following protocol presents an *in vitro* bioaccessibility extraction test for dioxin/furans in soil to be implemented at the Tronox Henderson, Nevada Site. The *in vitro* extraction method generally follows that reported in Ruby et al. (2002) and further reported upon in Finley et al. (2009). As an attachment to this protocol, and as requested by NDEP, the dioxin/furan congener profile for relevant Site soils samples is presented in Attachment 1 as it provides the basis for the identification of one dioxin/furan type at the Site.

Soil Sample Collection and Analysis

Soil sample collection will be targeted in areas of the Site where prior data indicate dioxin/furan TEQ concentrations generally between 1,000 and 3,000 ppt, and that include a representative range of organic carbon content, while also considering the site conceptual model. Soil samples (0-2 feet bgs in depth) will be collected from ten (10) different locations at the Site as follows:

Two samples from Area 4
Near RSAQ3
Near SA 84

Four samples from Area 2
Near SA 41
Near SA 114
Near SA 150
Near SA 167

Four samples from Area 1
Near SA 75

Near RSAH3
Near RSAL3
Near RSAK4

At each of these locations one additional sample will be collected and archived in the event that additional analysis is needed (10 for initial analysis and 10 archived samples). Samples will be shipped to Vista Laboratories for preparation and analysis. In the lab the soils will be air-dried and sieved to the <250 μm particle size samples. All samples will be analyzed for dioxin/furan content using isotope dilution gas chromatography-mass spectrometry, according to USEPA Method 1613, to ensure that the collected soils represent an appropriate dioxin/furan TEQ range for use in the bioaccessibility study. Samples will also be analyzed for organic carbon content.

Bioaccessibility Extraction Method

The extraction will be carried out in 1-L amber glass bottles with Teflon®-lined screw caps. The bottles will be partially immersed in a water bath to maintain a temperature of 37 C throughout the extraction procedure. Slow mixing will be provided by a stainless-steel paddle stirrer mounted in a rheostat-controlled motor (Arrow Engineering Model 1750®), or on a shaking water bath. A stirring/shaking rate of 30 rpm will be maintained during the *in vitro* extraction.

The test procedure involves extraction of 10 g of test soil (<250- μm size fraction) in 1 L of extraction fluid (1:100 soil:solution ratio), using a sequential extraction procedure that simulates a stomach phase (pH 1.5 with various enzymes, proteins, and fatty acids for 1 hour) followed by a small-intestinal phase (pH 7.2 with additional enzymes for 4 hours). Subsequent to the small-intestinal incubation, the extraction solution will be centrifuged (to remove any soil particles), and the extraction fluid will be submitted for analysis. The resultant data, in combination with the total concentrations of the target analyte(s) for each soil, will be used to calculate the fraction of chemical that is liberated from each test soil (i.e., fraction that is bioaccessible).

The extraction procedure will be conducted according to the following method:

- (Scale up stock solutions, below, as appropriate for the target number of samples to be extracted, including test soils and quality control samples.)
- Prepare 4 L of buffered stomach fluid by adding 60.06 g glycine (0.2 M; Sigma UltraPure®) to 4 L of Type II deionized (DI) water, and adjust to pH 1.5 with concentrated HCl (requires approx. 240 mL)



- Add 35.2 g of sodium chloride (NaCl, concentration of 150 mM in stomach fluid)
- Add 4.0 g of pepsin (activity of 800-2,500 units/mg, final concentration of 1.00 g/L in stomach fluid)
- Add 20 g bovine serum albumin (BSA; minimum 98 percent, final concentration of 5 g/L in stomach fluid)
- Add 10 g mucine (Type III, purified from porcine stomach; final concentration of 2.5 g/L in stomach fluid)
- Place 1 L of the stomach solution in each reaction vessel
- Add 6 mL of oleic acid (90%; Aldrich Chemical) to each extraction vessel
- Add 10 g of soil (<250 μm size fraction) to each reaction vessel
- Stir for 1 hour with paddle stirrer at 30 rpm to simulate stomach-phase extraction
- Bring reaction fluid in each vessel to pH 7.2 by adding sodium hydroxide (NaOH; 50 percent w/w, approximately 10 mL)
- Add 600 mg porcine pancreatin to each extraction vessel (activity equivalent to 8 x U.S.P. specifications)
- Add 4 g of bovine bile (50 percent bile acids, mixture of free and conjugated acids) to each extraction vessel
- Stir for 4 hours with paddle stirrer at 30 rpm
- After 4 hours of small-intestinal extraction time, allow the solids to settle, and decant all of the fluid from each reaction vessel into four 250-mL centrifuge tubes. Centrifuge at 3,000 Gs for 10 minutes and collect the supernatant in a 1-L amber glass bottle. Record the volume of the extraction fluid collected.
- Store all extracts at 4 C, and ship on ice to the analytical laboratory for analysis.

Mass Balance Testing

As a check on the recovery from the *in vitro* extraction, a mass balance test will be performed on the replicate extraction samples by adding the two following steps (outlined below) to the extraction protocol.



- Using DI water, wash the post-extraction soil from the reaction vessel onto a 1.0- μm glass-fiber filter. Wash any soil pellets in the centrifuge tubes onto the filter. Wash the filtered soil with 20 mL DI water. Add the filtrate to the extraction supernatant (in the 1-L amber glass bottle), and measure the volume of extraction fluid
- Collect the post-extraction soil, and remove 2 g for determination of percent moisture. Ship the remaining post-extraction soil (wet) to the analytical laboratory for analysis.

Quality Control

In addition to the 10 site soils, a set of samples will be included to allow for an assessment of data quality. These will include (at a minimum):

- Triplicate testing of one site soil
- Extraction blank
- Extraction spike (to be representative of the congener mix found in site soils)



ATTACHMENT 1: DIOXIN CONGENER PROFILE ANALYSIS

To evaluate the potential similarities or differences of dioxin/furan congener profiles from Site soils, a congener profile analysis was conducted for thirty-seven (37) soil samples collected from Areas I, II and IV. Most of these samples represent concentrations of dioxin/furan TEQ within the approximate range of 1,000 to 3,000 ppt, as this is the range of interest. However, for comparison purposes some additional congener profile for samples with higher reported dioxin/furnace concentrations are included and four (4) dioxin/furan samples collected near the former effluent pond berms SA 201-0.5B, RSAJ6-0.5B, RSAJ7-0.5B, RSAK3-0.5B) are included. Dioxin fingerprinting was not conducted for Area III samples, as all dioxin/furan TEQ results are below the BCL screening level of 1000 ppt.

Table 1 provides the dioxin/furan TEQ for the 37 and the individual congener results while Table 2 presents the congener data as a percentage of the total TEQ for the 37 samples. Total organic carbon content of the samples is also presented in Table 2. As shown in the first set of fingerprint profiles figures (Set 1), the congener fingerprints of all the samples are fairly consistent. With the exception of one sample (SA-129), dioxins generally account for less than 6% of the total TEQ and 2,3,7,8 TCDD constitutes a very minor component of each soil sample (less than 0.3%). In all samples, the hexa, hepta and octa furans account for the vast majority (at least 75%) of the total dioxin/furan TEQ in each sample, with OCDF accounting for at least 40% of the total TEQ concentration. This pattern is essentially identical to that reported by Finley et al., 2009 for a Magnesium facility in which chloride and metals were separated via an electrolytic process.

Table 3 provides the congener data as a percentage of the total TEQ for a set of supplemental soils samples in which dioxin/furan samples were collected at a depth of 1.0 – 1.5 feet bgs and 1.5-2.0 feet bgs. The second set of fingerprint profiles (Set 2) also shows that the fingerprint profiles for these samples are fairly consistent with depth.

Based on this evaluation, the dioxin/furan samples of interest have similar “fingerprints” and there is little variability of this pattern between samples collected in Areas I, II and IV, within high and low dioxin/furan TEQ concentrations or by depth. The fingerprint profiles will be confirmed based on the data collected as part of the bioaccessibility study.



Table 1 - Dioxin/Furan congener data for select soil samples ^{a)}

Area	Grid	sys_sample_code	2,3,7,8-TCDD (ng/kg)	1,2,3,7,8-PeCDD (ng/kg)	1,2,3,4,7,8-HxCDD (ng/kg)	1,2,3,7,8,9-HxCDD (ng/kg)	1,2,3,6,7,8-HxCDD (ng/kg)	1,2,3,4,6,7,8-HpCDD (ng/kg)	OCDD (ng/kg)	2,3,7,8-TCDF (ng/kg)	1,2,3,7,8-PeCDF (ng/kg)	2,3,4,7,8-PeCDF (ng/kg)	1,2,3,4,7,8-HxCDF (ng/kg)	1,2,3,7,8,9-HxCDF (ng/kg)	1,2,3,6,7,8-HxCDF (ng/kg)	2,3,4,6,7,8-HxCDF (ng/kg)	1,2,3,4,6,7,8-HpCDF (ng/kg)	1,2,3,4,7,8,9-HpCDF (ng/kg)	OCDF (ng/kg)	TEQ (ng/kg)
1 - Near Berm	I-3	SA201-0.5B	407	1630	1080	2560	2610	9370	8930	11400	19800	10000	60300	6390	38700	21300	135000	62300	363000	22,243
1 - Near Berm	J-6	RSAJ6-0.5B	990	1590	3440	7840	7090	20500	23000	29600	23000	11400	198000	22400	130000	69000	459000	210000	941000	60,611
1 - Near Berm	J-7	RSAJ7-0.5B	1020	2790	2020	4430	4040	15200	14600	27400	51200	24000	123000	12800	76000	9340	220000	91200	725000	41,935
1 - Near Berm	K-3	RSAK3-0.5B	1880	2880	2680	5840	5290	17000	22700	11700	40700	20200	122000	15200	79300	43400	315000	148000	821000	45,635
1	H-3	RSAH3-0.5B	21.7	99.1	75.4	158	137	462	491	601	1780	739	3630	281	2480	1150	7600	2740	16000	1,360
1	J-2	RSAJ2-0.5B	156	485	350	559	461	1360	1670	1860	3030	1240	10200	913	6740	1440	20800	10500	52900	3,699
1	J-5	RSAJ5-0.5B	118	389	251	675	602	1450	1860	1700	2870	1230	10400	439	6130	1570	17100	7600	52900	3,417
1	J-6	SA127-0.5B	32.6	85.4	56.6	129	105	334	364	615	1420	560	3870	307	2460	333	7270	3460	16100	1,232
1	K-4	RSAK4-0.5B	32.1	118	100	221	178	514	527	1430	1300	581	4410	296	3060	734	10000	3840	21300	1,556
1	K-8	RSAK8-0.5B	38.5	117	91.7	194	153	791	686	2240	1620	692	5450	361	3440	424	10900	5350	26900	1,826
1	L-3	RSAL3-0.5B	23.9	83.8	67.4	152	138	534	917	1100	1330	388	4150	179	1920	286	4680	2170	12500	1,141
1	L-4	SA189-0.5B	25.9	73.5	59.2	120	103	369	413	2000	1140	499	2770	164	1840	318	6170	2600	16500	1,117
1	L-7	SA75-0.5B	24.3	91.3	67.8	145	127	380	448	1970	1750	897	2720	222	1580	797	3960	1700	13200	1,265
2 - Ditch	M-7	SA155-0.5B	139	482	408	844	766	2490	2770	2430	6110	2990	16800	1390	10700	5790	29800	13400	85000	6,097
2	L-5	SA167-0.5B	41.5	151	143	271	264	850	1040	621	1960	882	6930	432	4100	1140	7580	3100	18800	2,027
2	M-5	SA104-0.5B	51	195	143	248	227	742	1250	2650	1970	934	4950	473	2810	1640	8600	4150	35700	2,045
2	M-5	SA129-0.5B	22.6	68	282	212	141	633	5390	4850	1480	1050	2290	60.9	677	587	2780	1530	30700	1,420
2	M-6	SA175-0.5B	159	429	414	698	626	1850	2510	6540	4350	2050	13100	1330	7130	4170	25700	11700	86500	5,154
2	M-7	SA63-0.5B	155	493	480	871	763	2310	3290	6100	4710	2350	15500	1550	8450	4840	31000	13700	111000	5,854
2	M-7	SA86-0.5B	185	619	528	835	786	2790	3860	3320	7250	4130	14400	1370	7560	4420	24900	11000	75100	5,993
2	M-7	SA92-0.5B	18.8	89.7	53.9	130	112	377	377	1690	1460	506	3460	266	2210	1230	6410	3050	17600	1,323
2	N-5	SA58-0.5B	28.2	67.5	55.8	119	96.4	376	385	1750	1380	713	3650	372	2390	1480	5360	2660	18700	1,432
2	N-6	SA105-0.5B	50	84.3	54.9	140	118	422	456	2050	1390	682	3320	391	2090	1200	5000	2490	17900	1,402
2	N-6	SA150-0.5B	76.8	283	227	448	367	965	1450	4170	2430	1170	7620	722	4330	2530	14000	6190	50800	3,052
2	N-6	SA60-0.5B	101	304	216	353	366	1370	2810	6030	4030	2120	11400	1380	6500	4020	21500	10700	85100	4,550
2	N-7	SA49-0.5B	114	383	271	546	445	1270	1970	4290	4090	2020	9070	1140	5930	3510	16100	8130	56000	4,018
2	N-7	SA107-0.5B	67.7	227	143	333	282	1080	757	4240	3830	1800	7780	837	4770	2670	11400	5310	32200	3,243
2	O-5	SA114-0.5B	73.5	263	213	417	356	1250	1780	881	2150	1150	6930	651	3710	2210	14600	6710	43200	2,522
2	O-5	SA187-0.5B	64.6	254	192	386	374	1060	1470	1060	2060	1140	6380	448	3570	1890	9410	4080	39200	2,310
2	O-5	SA41-0.5B	50.1	167	112	239	200	740	875	1180	2000	918	6460	574	3890	2070	13700	5420	43000	2,237
2	O-6	SA200-0.5B	13.9	52.1	42.8	88.1	77.3	297	376	1100	816	366	2940	212	1730	1030	6470	3060	22700	1,027
2	O-6	SA51-0.5B	24.1	78.9	53.8	121	105	407	813	1410	972	508	3210	244	1890	1090	6040	3050	20600	1,198
4	Q-3	RSAQ3-0.5B	720	2200	1000	1100	1400	5600	5200	12000	19000	7800	30000	2100	15000	2300	47000	19000	140000	13,000
4	Q-3	RSAQ3009-0.5B (dup)	230	760	530	1000	880	3300	3500	3700	6900	2300	15000	1700	8400	1900	28000	16000	110000	5,700
4	Q-3	SA169-0.5B	79	210	140	220	240	870	820	1400	2300	1200	4700	450	2900	680	9900	4600	30000	2,000
4	Q-4	SA84-0.5B	22	64	59	130	100	450	690	770	1400	650	3500	390	2000	540	9100	3800	31000	1,200
4	Q-5	SA156-0.5B	74	150	76	150	140	480	750	2100	1200	820	2500	170	1600	400	5500	2100	15000	1,300

Note: All Area 4 data are unvalidated

^{a)} For use in Set 1 profiles

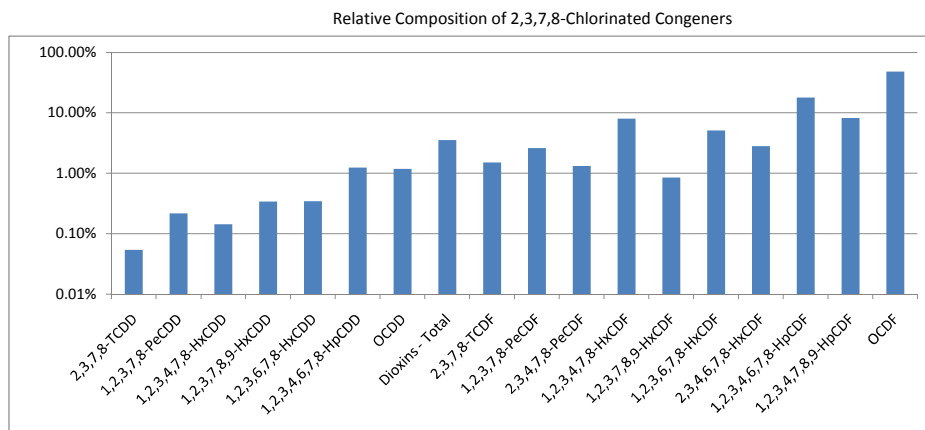
Table 3 - Dioxin/Furan congener percentage of TEQ for select supplemental (depth) soil samples ^{a)}

	Start Depth	Area	Grid	2,3,7,8-TCC	1,2,3,7,8-PC	1,2,3,4,7,8-PC	1,2,3,7,8,9-PC	1,2,3,6,7,8-PC	1,2,3,4,6,7,OCDD	2,3,7,8-TCC	1,2,3,7,8-PC	1,2,3,4,7,8-PC	1,2,3,4,7,8-PC	1,2,3,7,8,9-PC	1,2,3,6,7,8-PC	2,3,4,6,7,8-PC	1,2,3,4,6,7,OCDF	1,2,3,4,7,8,OCDF			
SA201-0.5B	0.5 ft	1 - Near Be	I-3	0.05%	0.22%	0.14%	0.34%	0.35%	1.24%	1.18%	1.51%	2.62%	1.32%	7.99%	0.85%	5.13%	2.82%	17.89%	8.25%	48.09%	
SA201-1BR	1 ft	1 - Near Be	I-3	0.09%	0.33%	0.24%	0.47%	0.61%	1.94%	1.67%	1.84%	3.92%	1.96%	8.83%	0.35%	5.89%	1.24%	15.50%	8.04%	47.08%	100.00%
SA201-1.5BR	1.5 ft	1 - Near Be	I-3	0.05%	0.19%	0.11%	0.22%	0.27%	1.29%	1.02%	1.14%	3.31%	1.43%	8.20%	1.07%	5.99%	1.28%	20.50%	9.78%	44.15%	100.00%
RSAH3-0.5B	0.5 ft	1	H-3	0.06%	0.26%	0.20%	0.41%	0.36%	1.20%	1.28%	1.56%	4.63%	1.92%	9.44%	0.73%	6.45%	2.99%	19.77%	7.13%	41.62%	
RSAH3-1BR	1 ft	1	H-3	0.08%	0.27%	0.17%	0.39%	0.37%	1.25%	1.26%	2.02%	4.38%	2.02%	7.91%	1.03%	5.55%	1.13%	16.83%	6.56%	48.80%	100.00%
RSAH3-1.5BR	1.5 ft	1	H-3	0.07%	0.27%	0.16%	0.33%	0.34%	1.19%	1.11%	2.05%	4.52%	2.13%	7.94%	1.02%	5.63%	1.19%	17.07%	7.17%	47.79%	100.00%
RSAL3-0.5B	0.5 ft	1	L-3	0.08%	0.27%	0.22%	0.50%	0.45%	1.74%	2.99%	3.59%	4.34%	1.27%	13.55%	0.58%	6.27%	0.93%	15.28%	7.09%	40.82%	
RSAL3-1BR	1 ft	1	L-3	0.11%	0.50%	0.29%	0.18%	1.08%	3.79%	3.21%	3.50%	6.13%	2.48%	9.92%	0.58%	5.54%	1.14%	15.17%	5.54%	40.84%	100.00%
RSAL3-1.5BR	1.5 ft	1	L-3	0.34%	0.38%	0.25%	0.92%	0.53%	2.26%	3.39%	3.77%	5.47%	2.45%	8.30%	0.85%	4.71%	1.13%	14.71%	7.17%	43.37%	100.00%
SA155-0.5B	0.5 ft	2 - Ditch	M-7	0.08%	0.26%	0.22%	0.46%	0.42%	1.37%	1.52%	1.33%	3.35%	1.64%	9.22%	0.76%	5.87%	3.18%	16.35%	7.35%	46.62%	
SA155-1BR	1 ft	2 - Ditch	M-7	0.08%	0.30%	0.25%	0.45%	0.45%	1.40%	1.03%	1.92%	4.02%	2.20%	8.42%	1.31%	5.14%	1.03%	17.77%	9.35%	44.88%	100.00%
SA155-1.5BR	1.5 ft	2 - Ditch	M-7	0.08%	0.15%	0.19%	0.31%	0.44%	1.46%	1.90%	1.68%	3.45%	1.86%	6.20%	0.71%	5.31%	0.97%	19.48%	7.08%	48.70%	100.00%
SA104-0.5B	0.5 ft	2	M-5	0.08%	0.29%	0.21%	0.37%	0.34%	1.11%	1.87%	3.97%	2.95%	1.40%	7.42%	0.71%	4.21%	2.46%	12.89%	6.22%	53.50%	
SA104-1BR	1 ft	2	M-5	0.10%	0.21%	0.12%	0.25%	0.34%	1.17%	1.64%	2.38%	3.57%	2.08%	6.24%	0.65%	5.05%	1.26%	19.33%	6.54%	49.06%	100.00%
SA104-1.5BR	1.5 ft	2	M-5	0.28%	0.27%	0.24%	0.17%	0.42%	1.67%	8.67%	2.17%	2.00%	0.27%	6.17%	0.85%	4.50%	1.00%	16.67%	8.00%	46.66%	100.00%
SA200-0.5B	0.5 ft	2	O-6	0.03%	0.13%	0.10%	0.21%	0.19%	0.72%	0.91%	2.66%	1.97%	0.88%	7.11%	0.51%	4.18%	2.49%	15.64%	7.40%	54.87%	
SA200-1BR	1 ft	2	O-6	0.05%	0.20%	0.13%	0.23%	0.27%	1.07%	1.12%	1.47%	3.56%	1.88%	8.14%	0.71%	4.68%	1.22%	16.27%	8.14%	50.85%	100.00%
SA200-1.5BR	1.5 ft	2	O-6	0.06%	0.20%	0.08%	0.23%	0.36%	1.01%	1.09%	5.79%	5.07%	3.62%	7.97%	0.47%	4.71%	1.63%	15.93%	8.33%	43.45%	100.00%

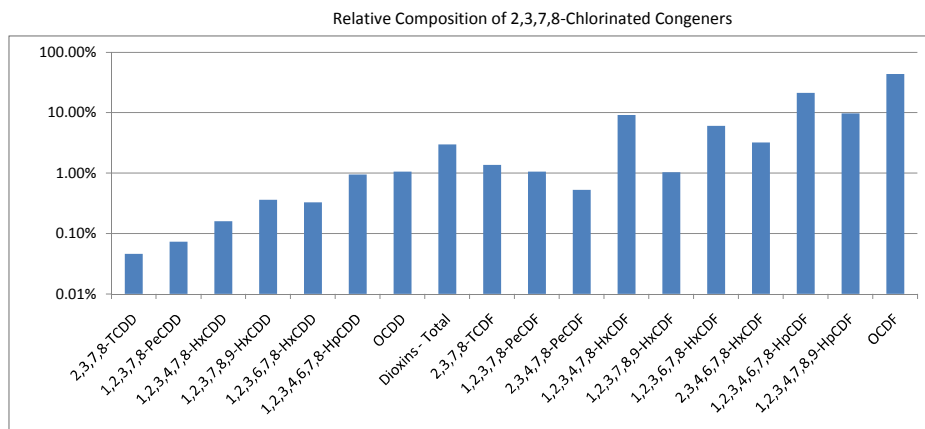
^{a)} For use in Set 2 profiles

Set 1 - Dioxin/Furan profiles for select soil samples

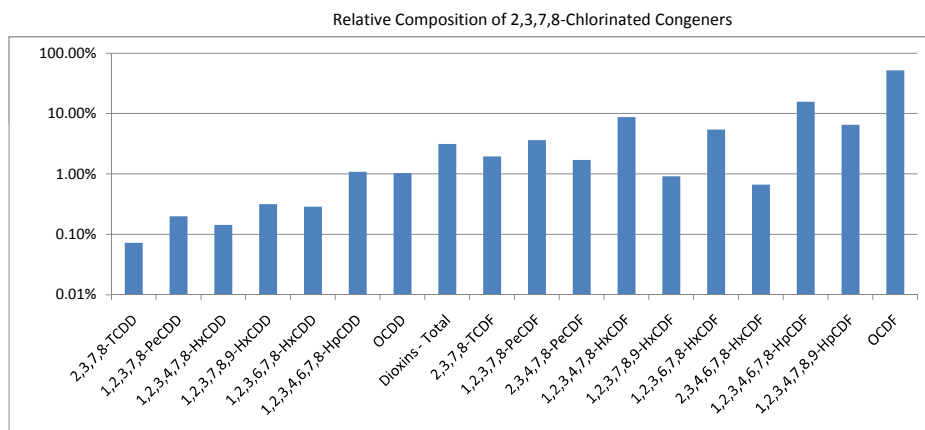
Area 1 - Near Berm
 Grid I-3
 Sample SA201-0.5B
 TTEQ (ppt) 22,243



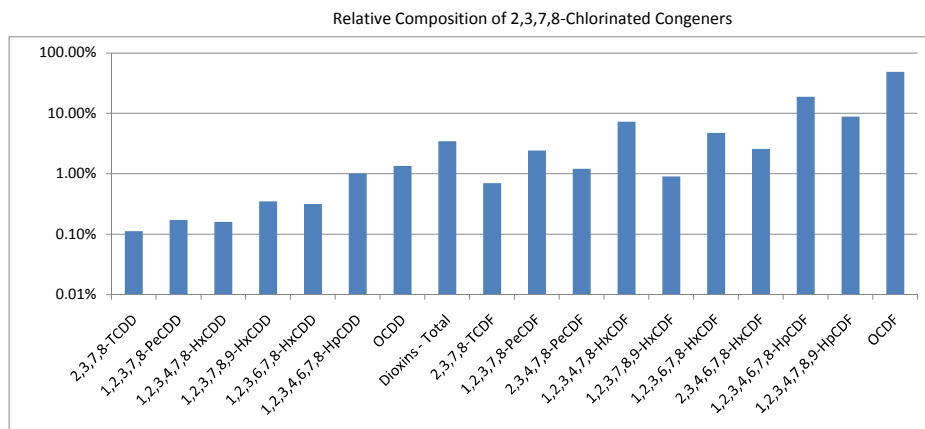
Area 1 - Near Berm
 Grid J-6
 Sample RSAJ6-0.5B
 TTEQ (ppt) 60,611



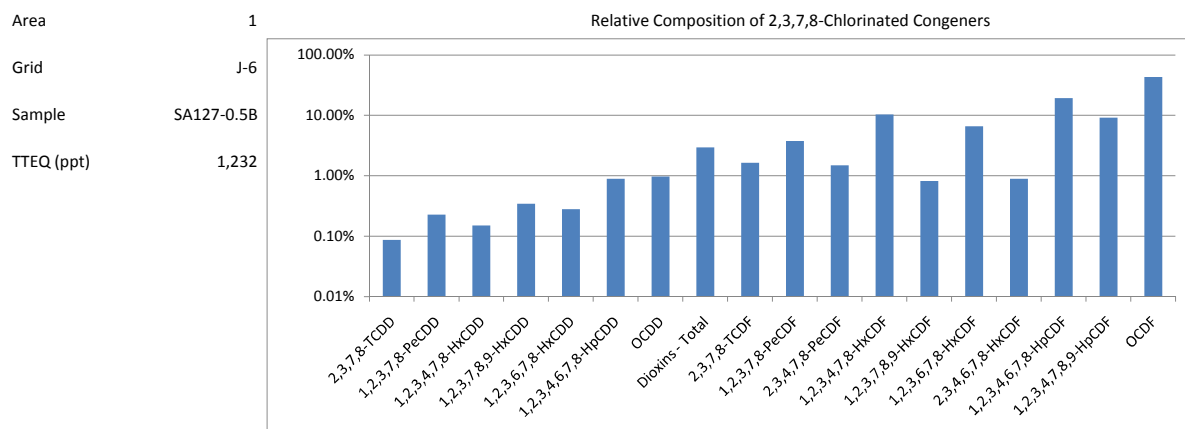
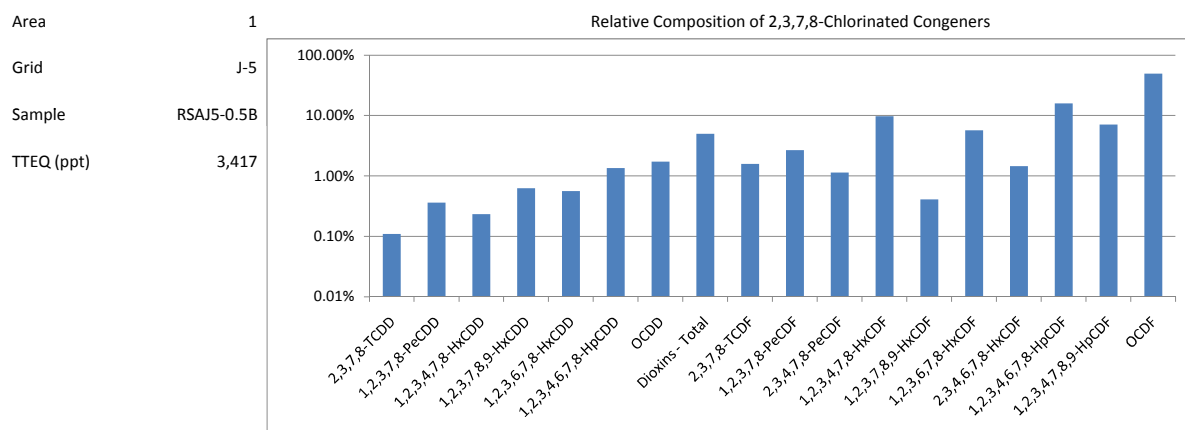
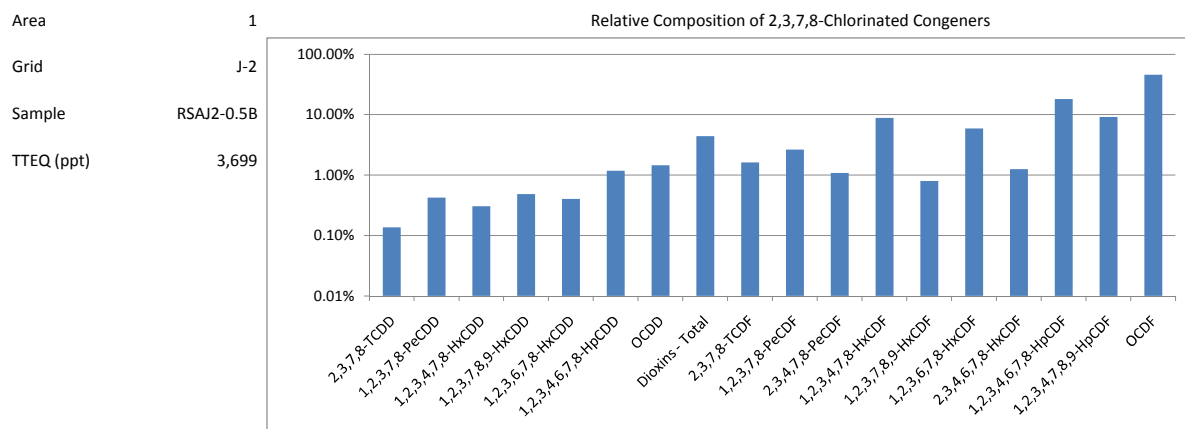
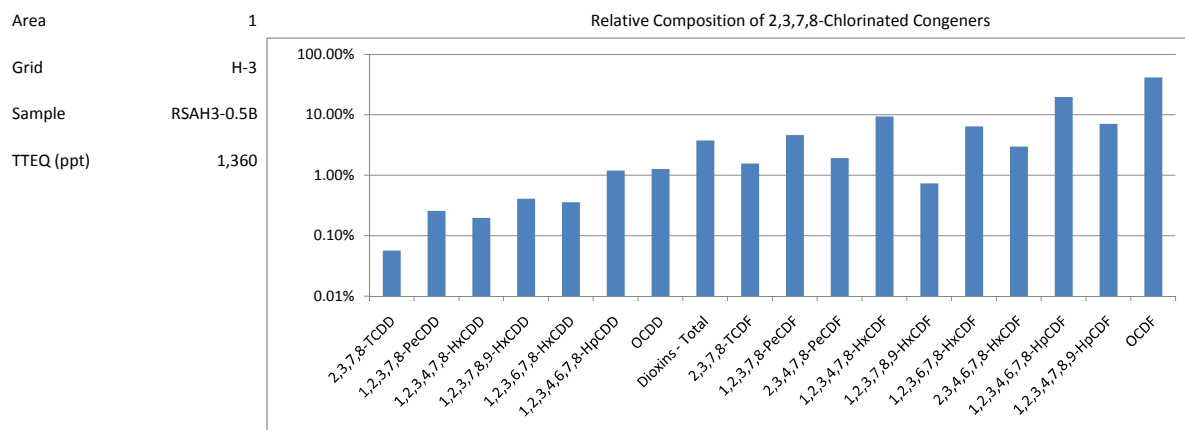
Area 1 - Near Berm
 Grid J-7
 Sample RSAJ7-0.5B
 TTEQ (ppt) 41,935



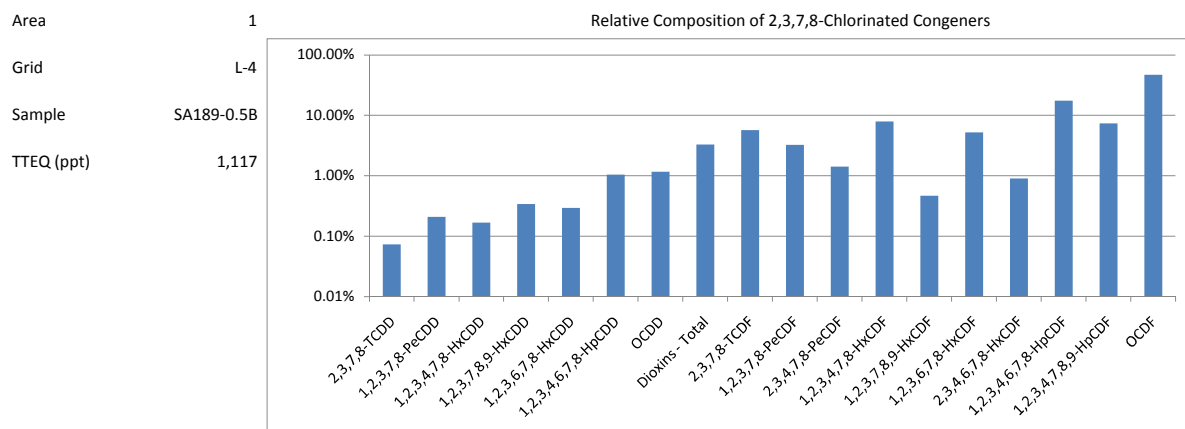
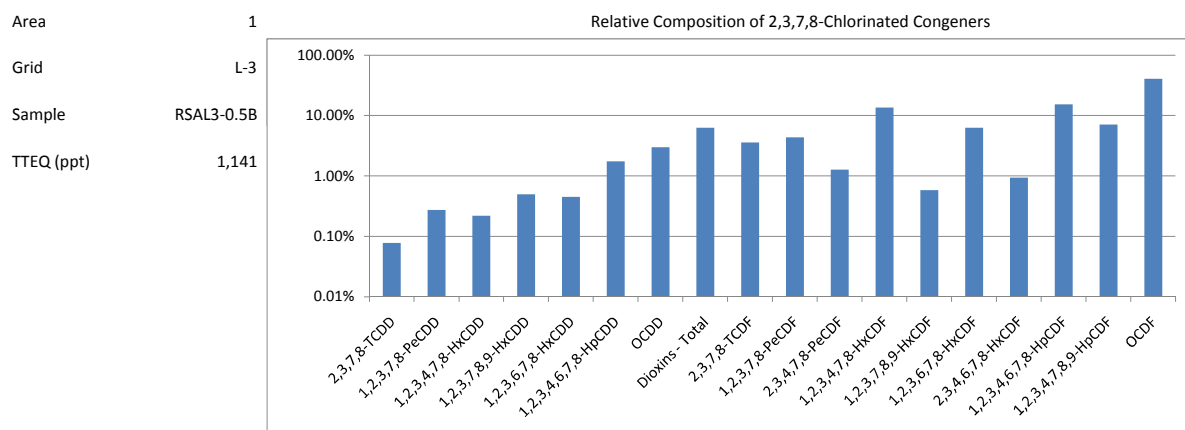
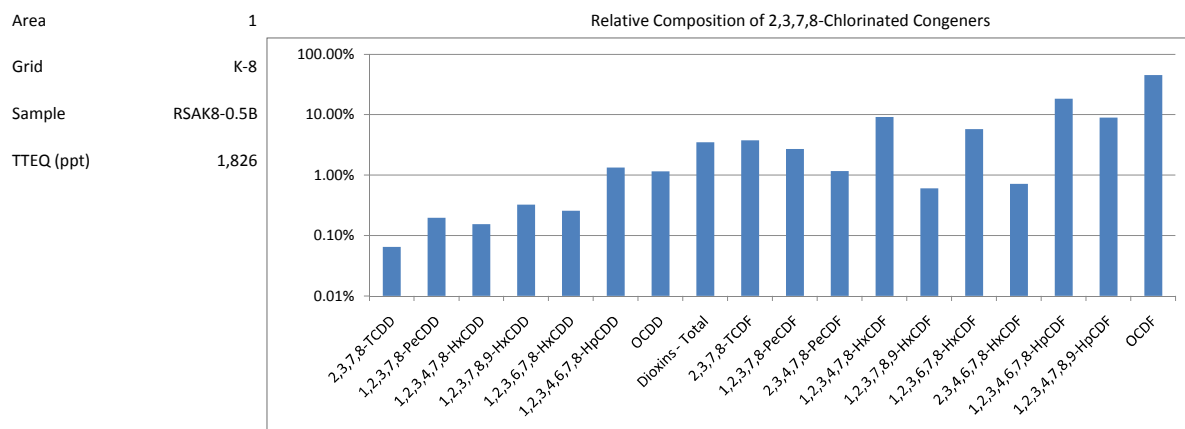
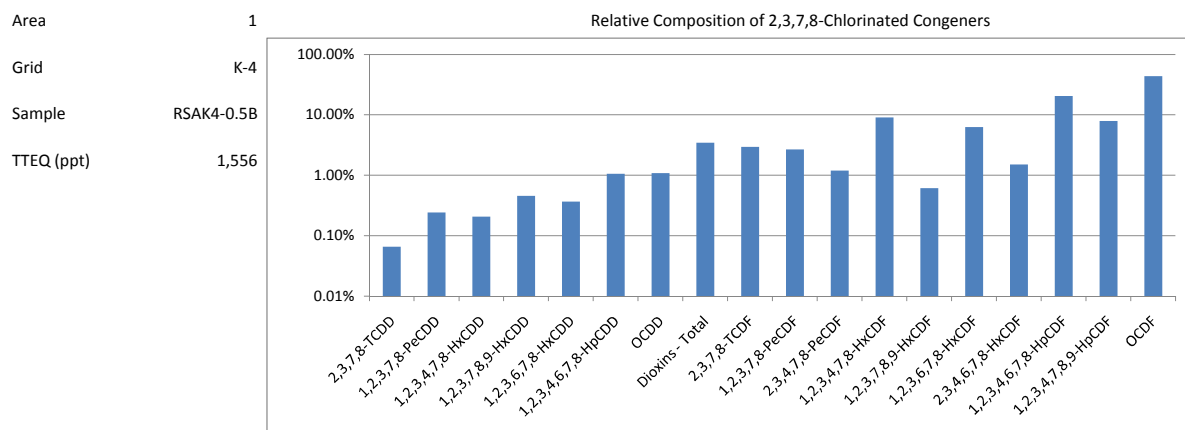
Area 1 - Near Berm
 Grid K-3
 Sample RSAK3-0.5B
 TTEQ (ppt) 45,635



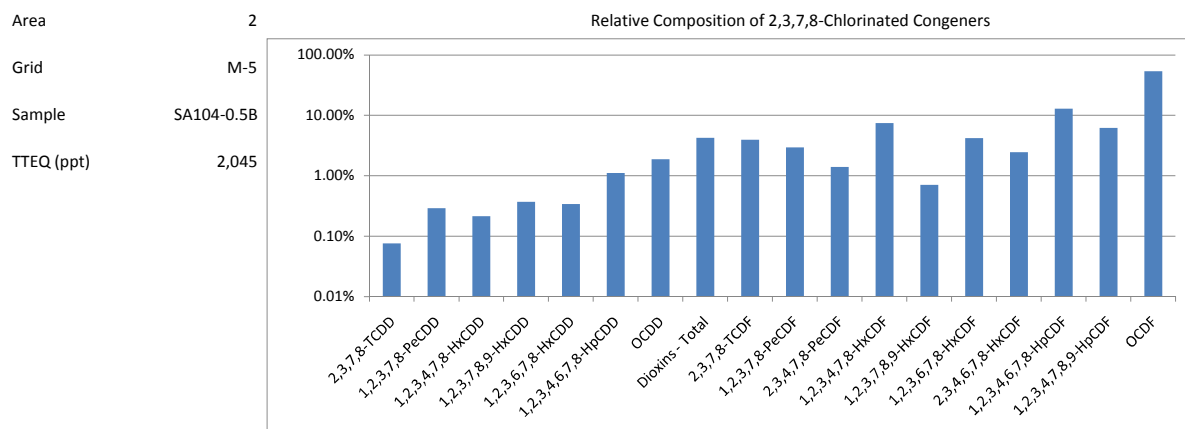
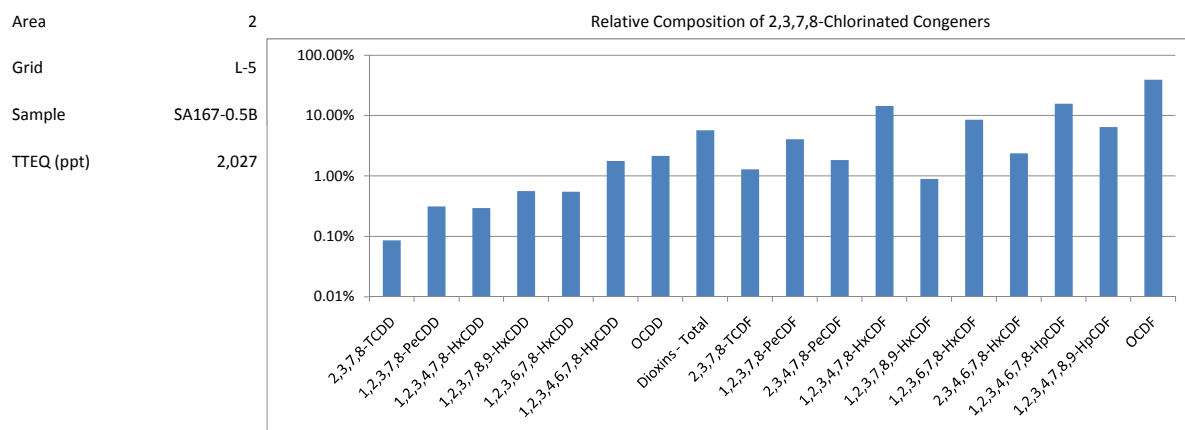
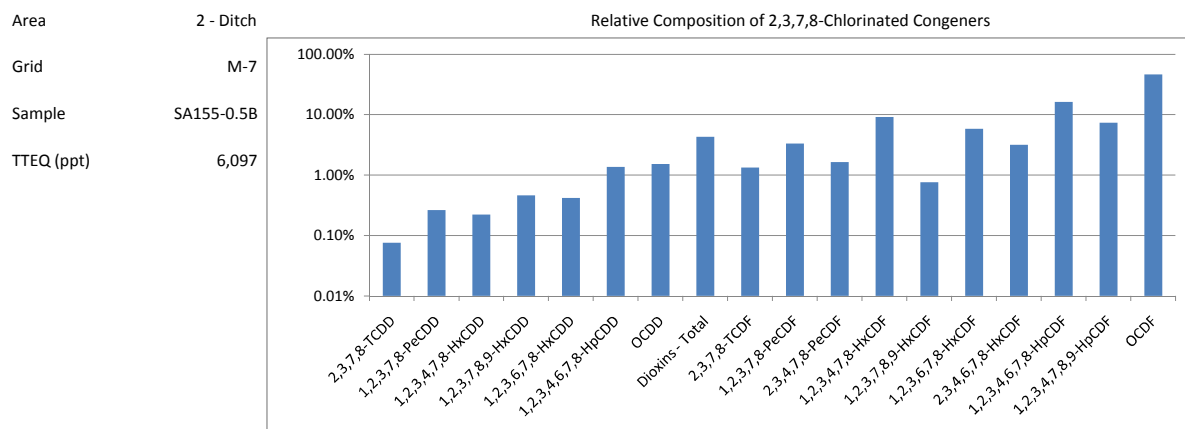
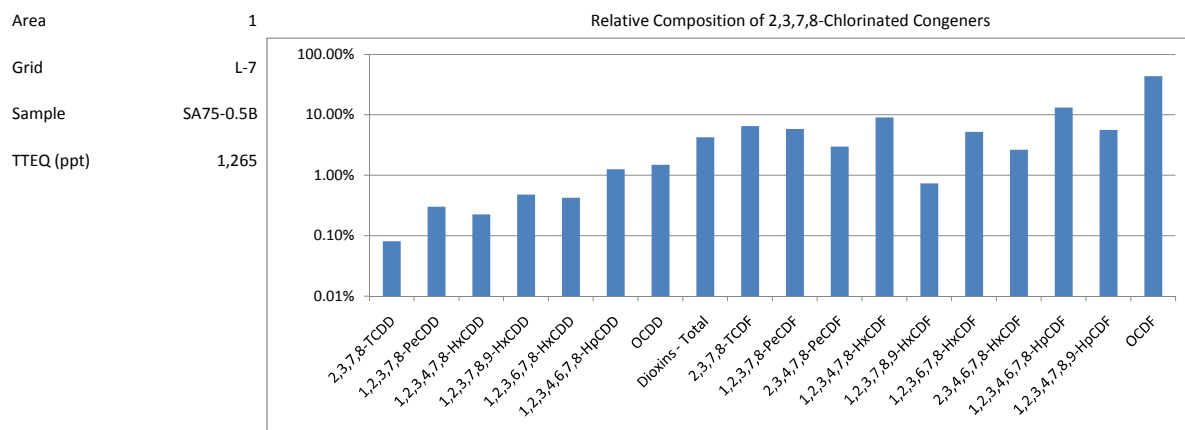
Set 1 - Dioxin/Furan profiles for select soil samples



Set 1 - Dioxin/Furan profiles for select soil samples

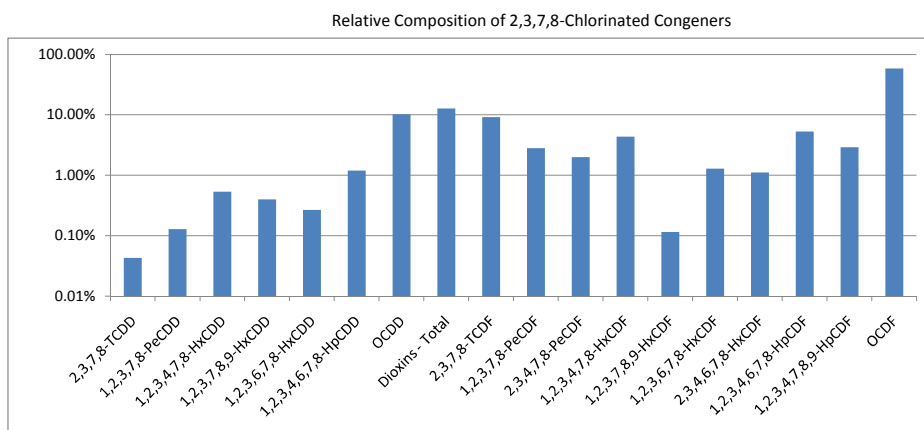


Set 1 - Dioxin/Furan profiles for select soil samples

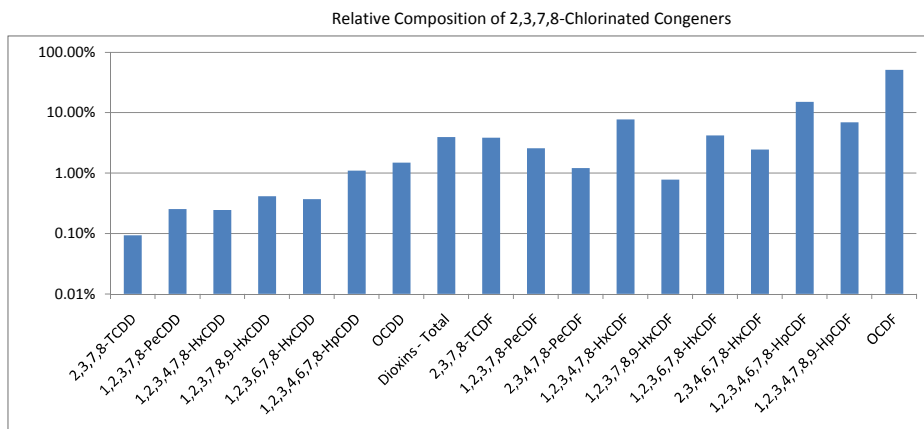


Set 1 - Dioxin/Furan profiles for select soil samples

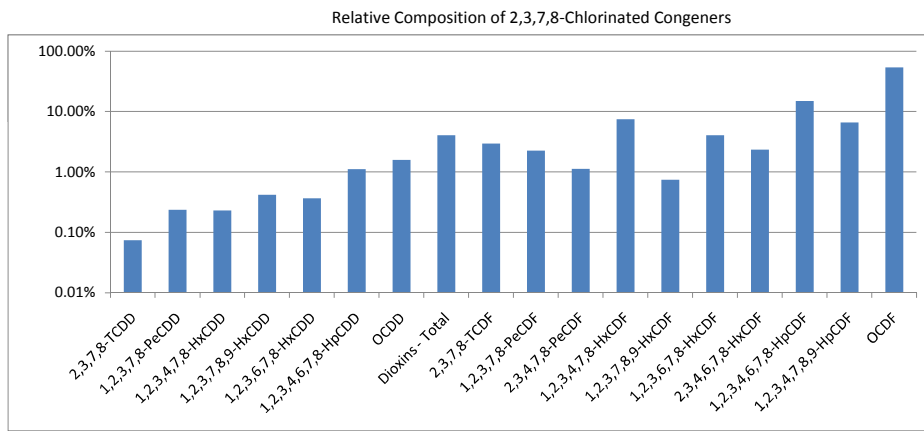
Area 2
 Grid M-5
 Sample SA129-0.5B
 TTEQ (ppt) 1,420



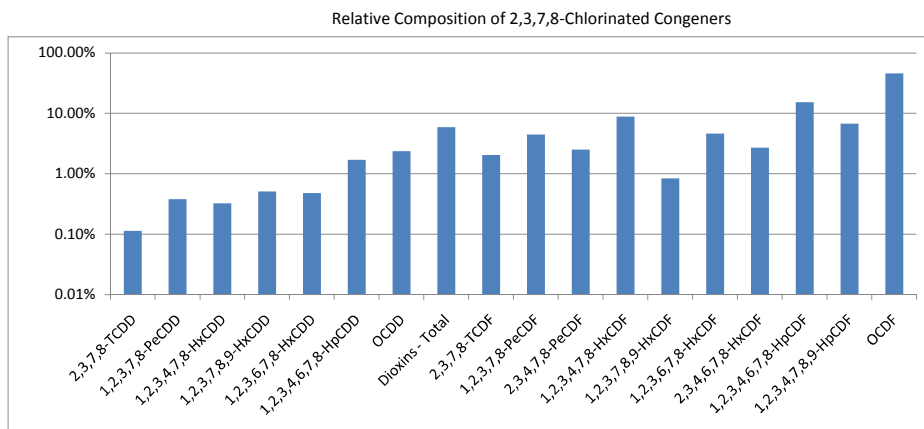
Area 2
 Grid M-6
 Sample SA175-0.5B
 TTEQ (ppt) 5,154



Area 2
 Grid M-7
 Sample SA63-0.5B
 TTEQ (ppt) 5,854

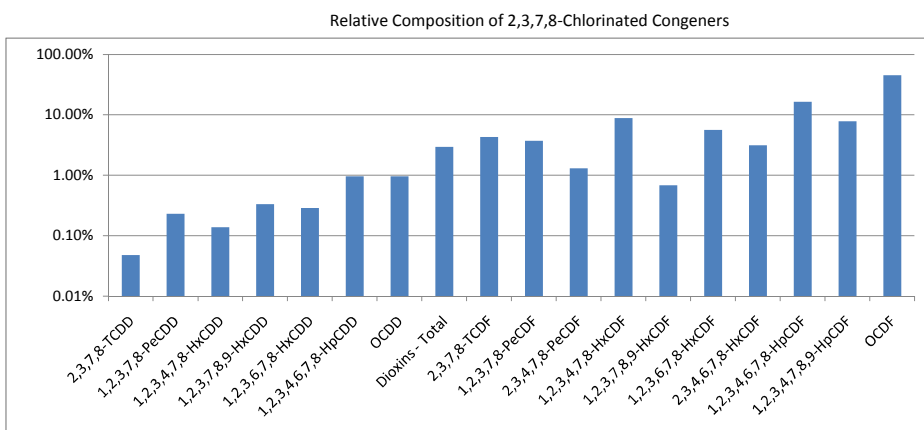


Area 2
 Grid M-7
 Sample SA86-0.5B
 TTEQ (ppt) 5,993

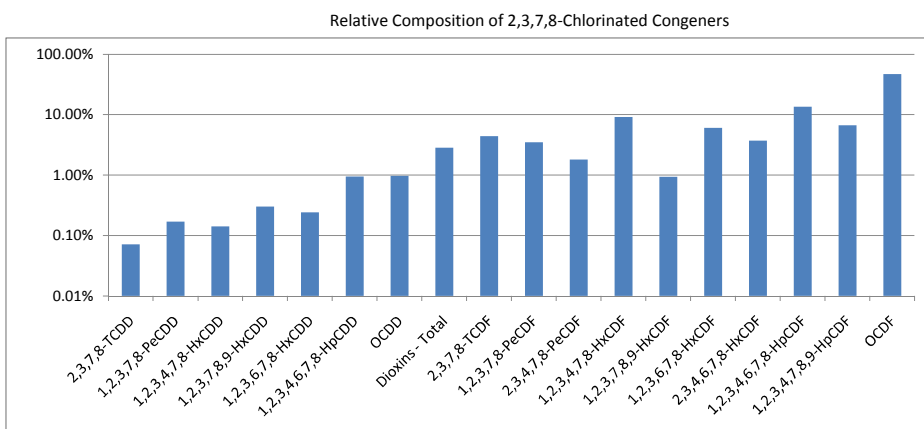


Set 1 - Dioxin/Furan profiles for select soil samples

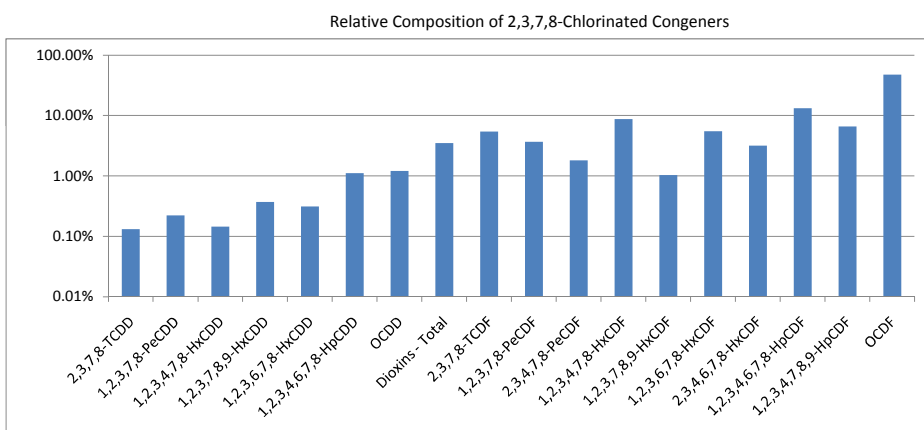
Area 2
 Grid M-7
 Sample SA92-0.5B
 TTEQ (ppt) 1,323



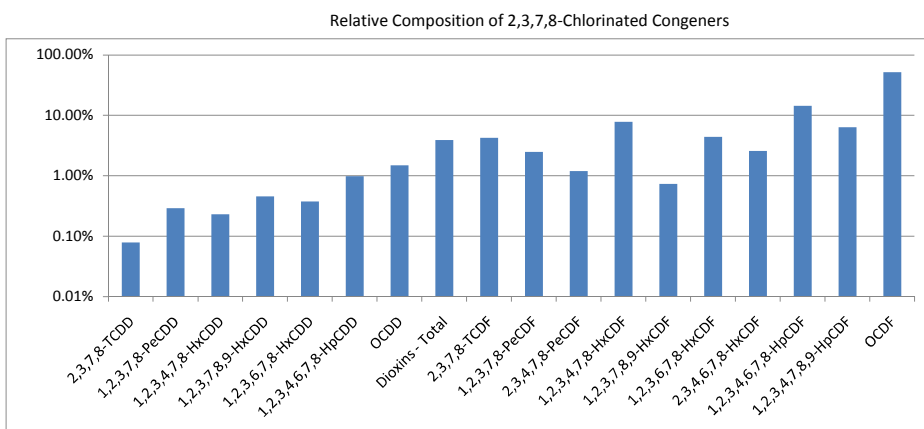
Area 2
 Grid N-5
 Sample SA58-0.5B
 TTEQ (ppt) 1,432



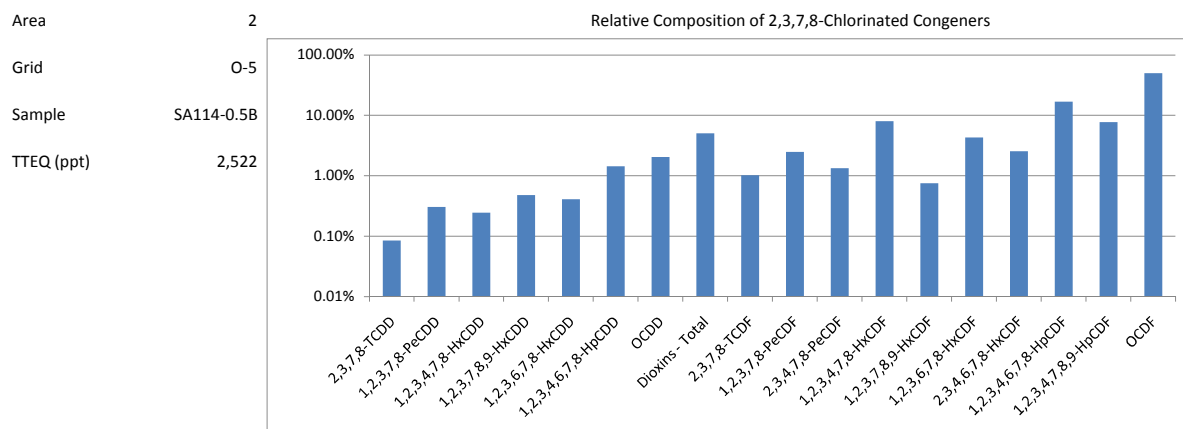
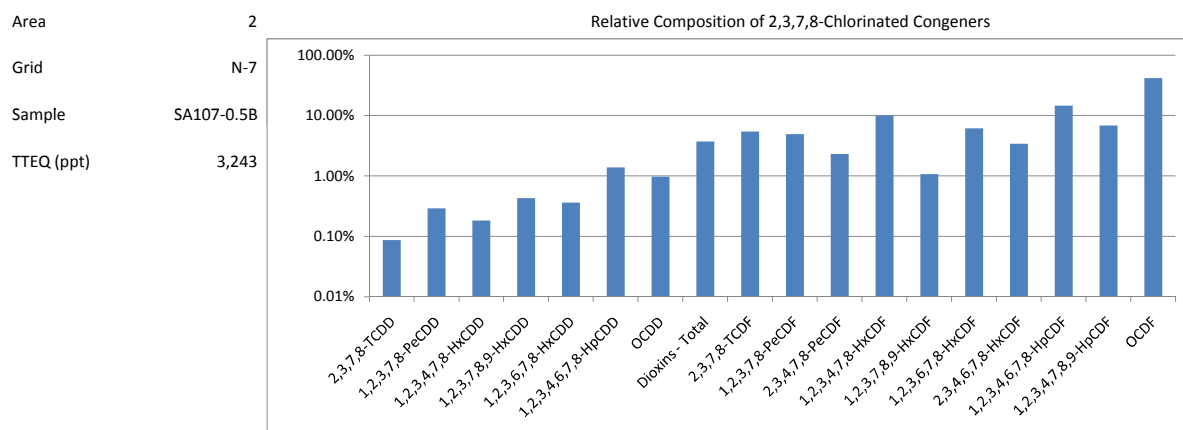
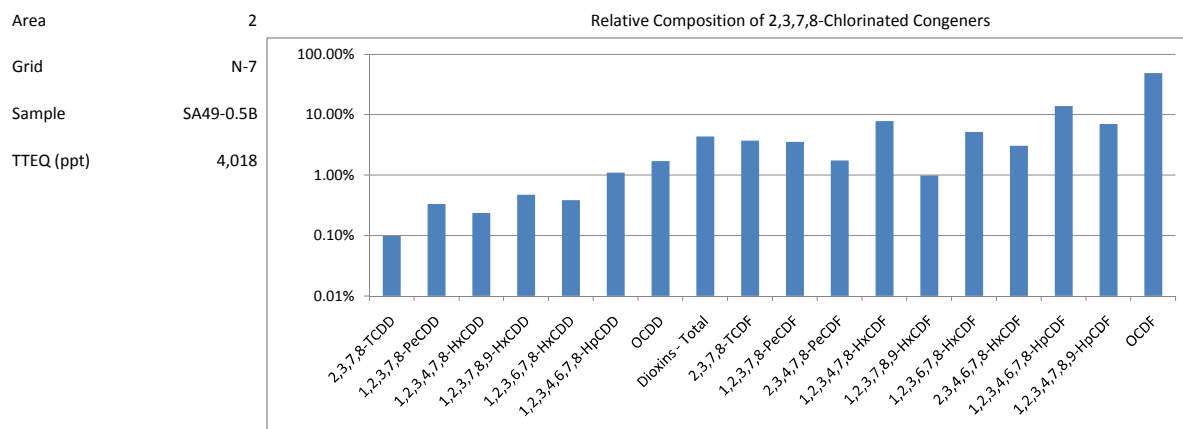
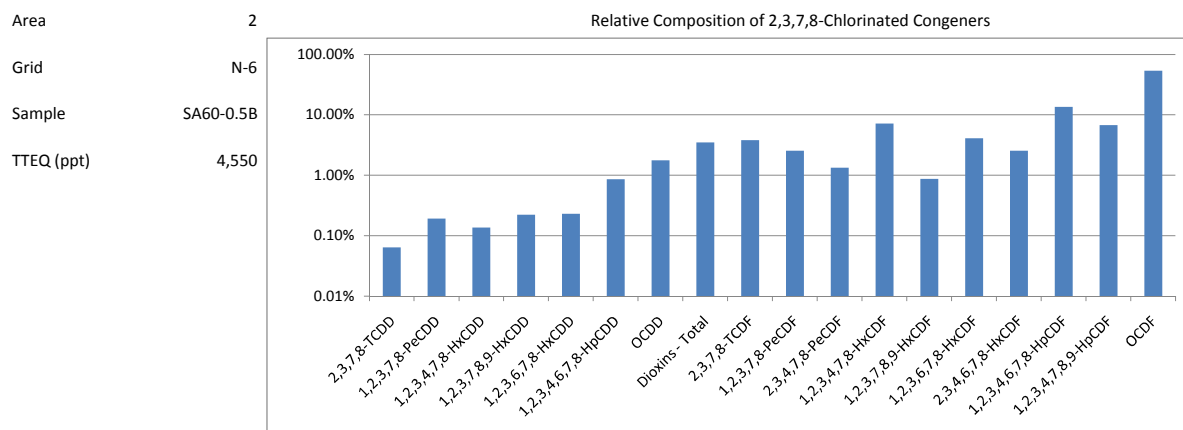
Area 2
 Grid N-6
 Sample SA105-0.5B
 TTEQ (ppt) 1,402



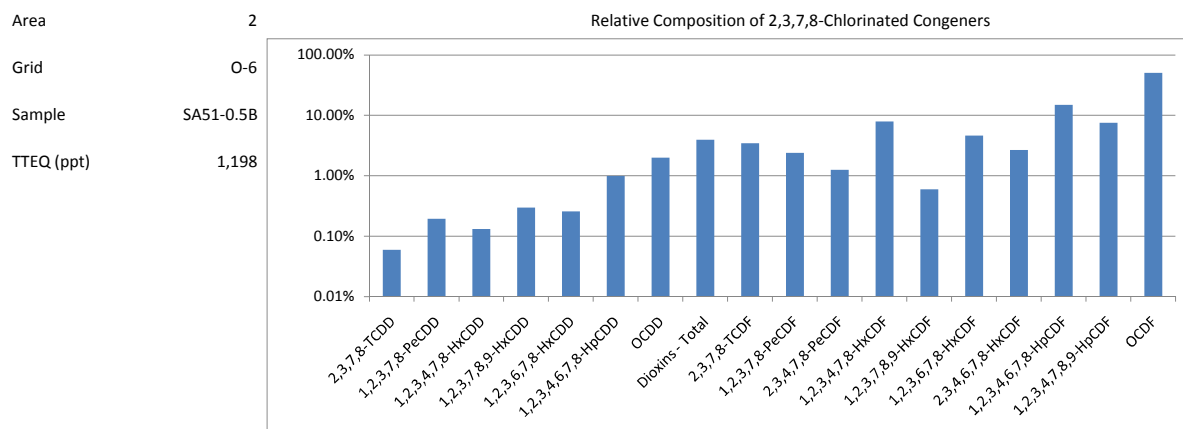
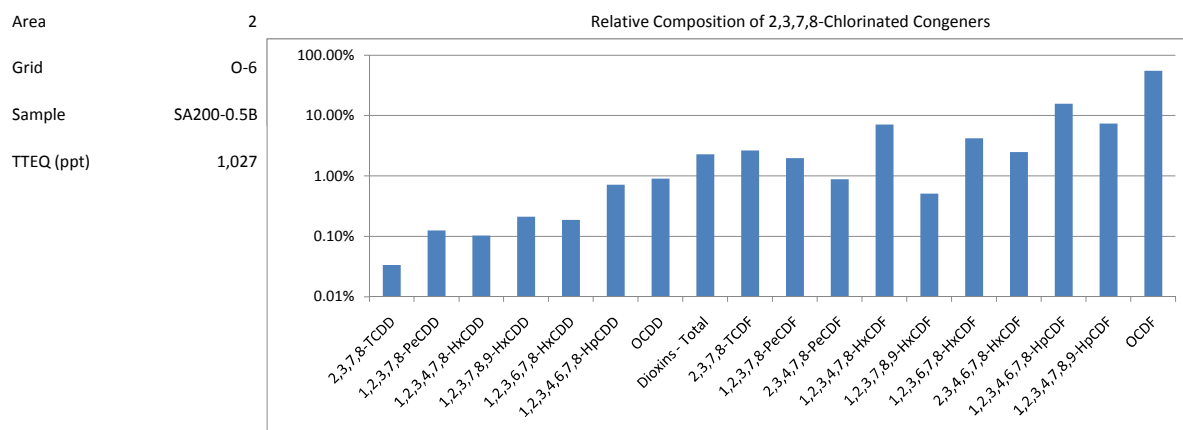
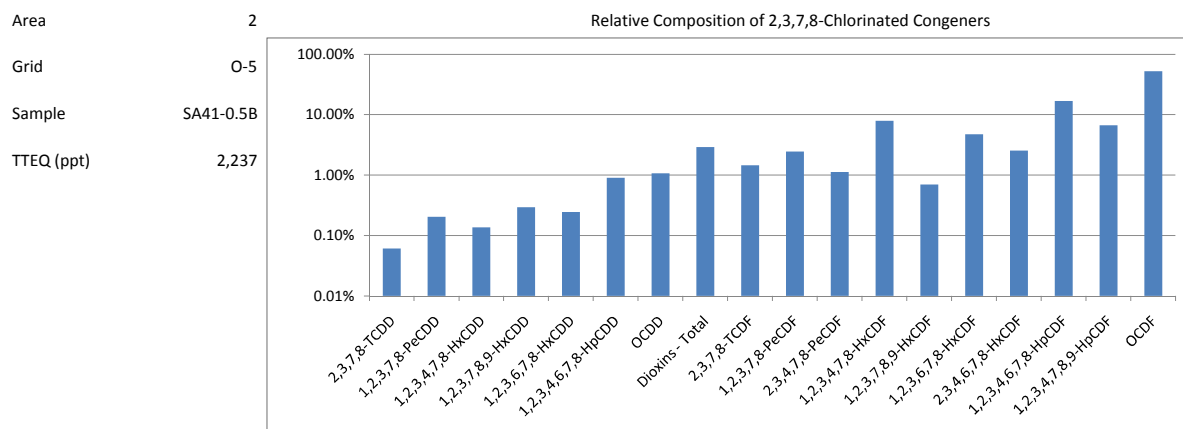
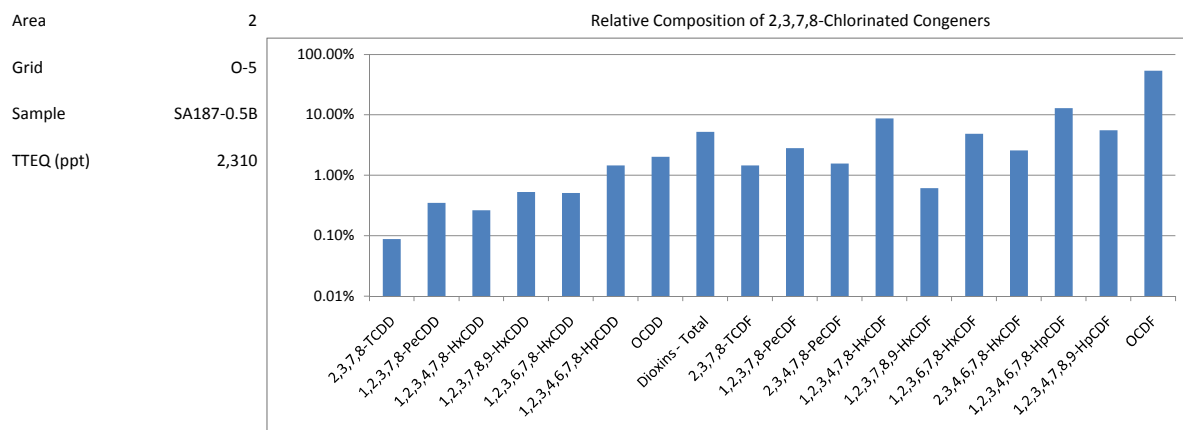
Area 2
 Grid N-6
 Sample SA150-0.5B
 TTEQ (ppt) 3,052



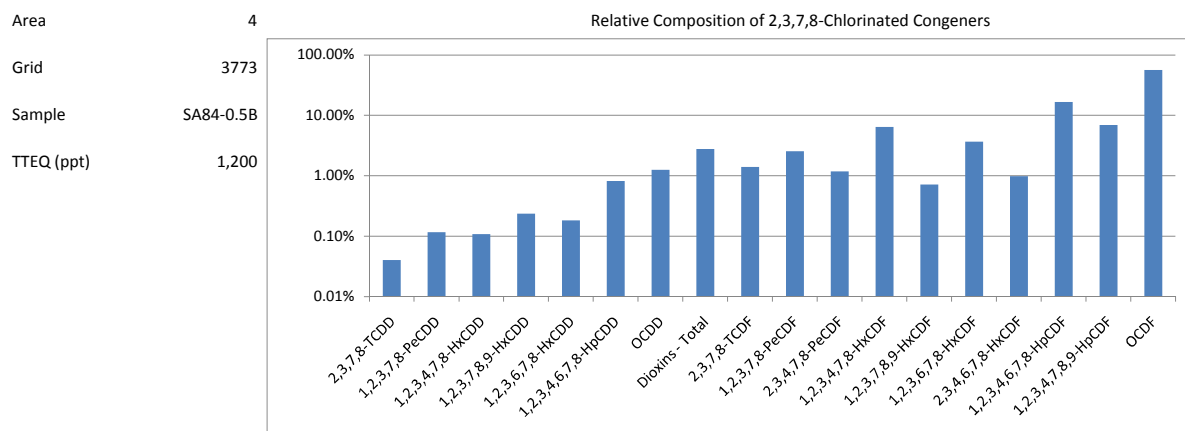
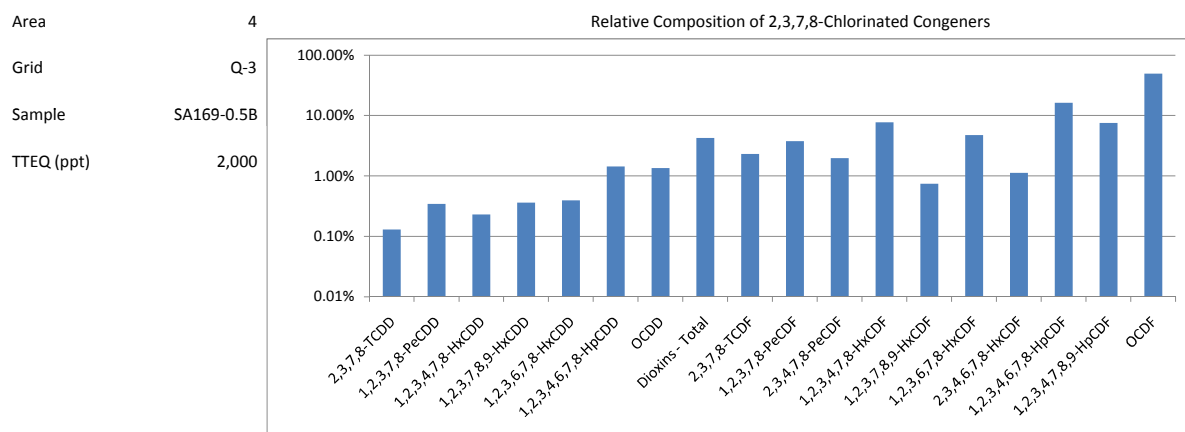
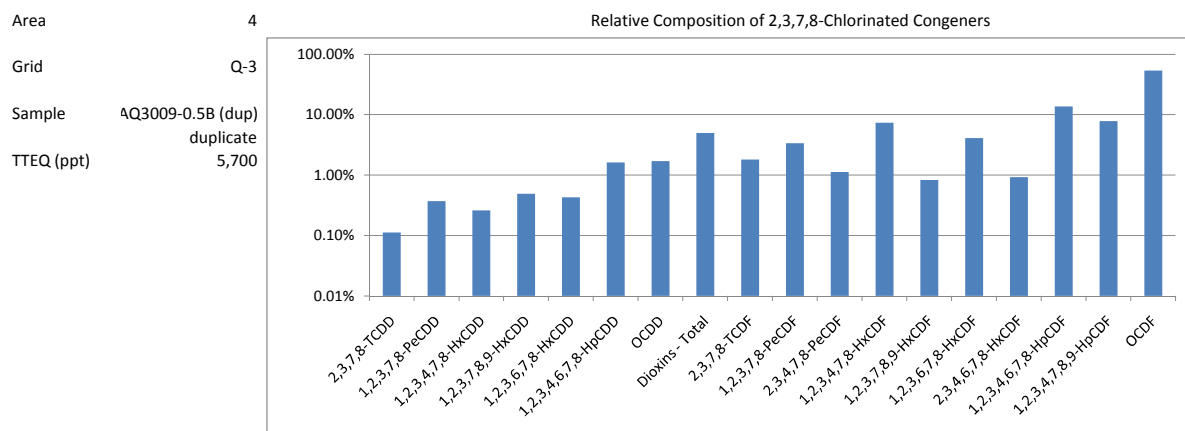
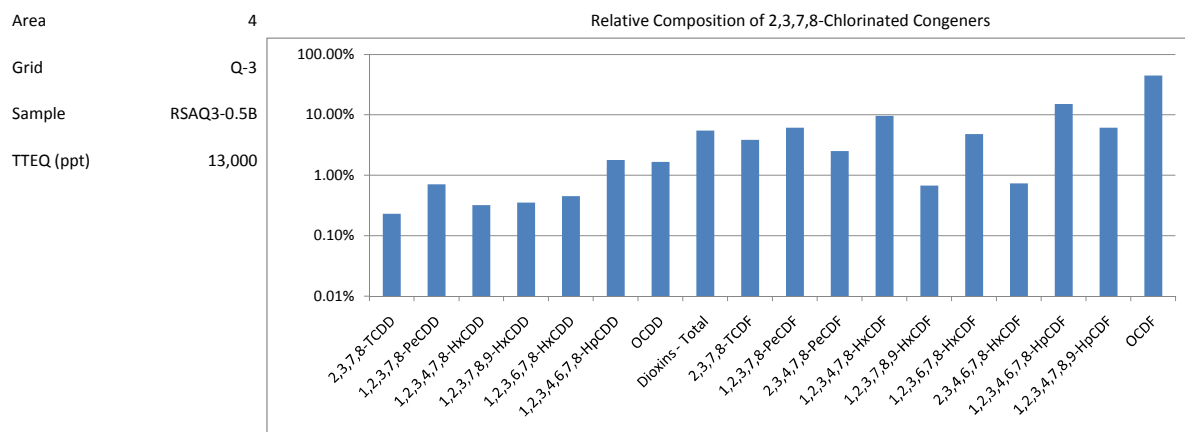
Set 1 - Dioxin/Furan profiles for select soil samples



Set 1 - Dioxin/Furan profiles for select soil samples

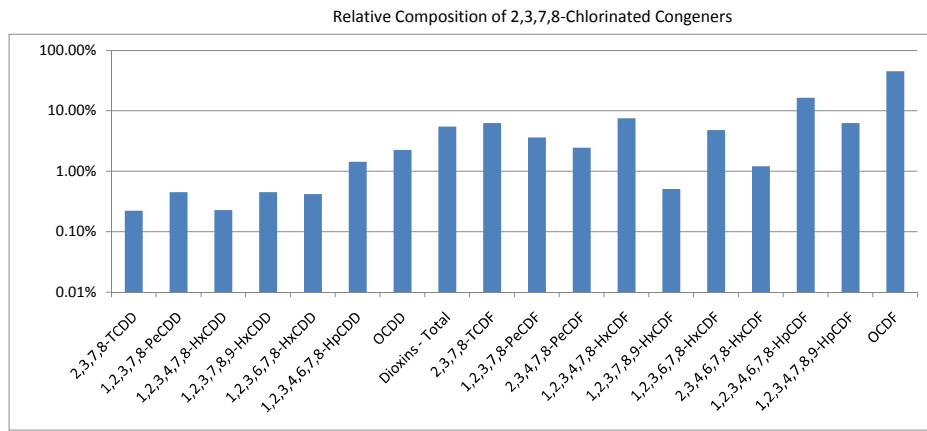


Set 1 - Dioxin/Furan profiles for select soil samples



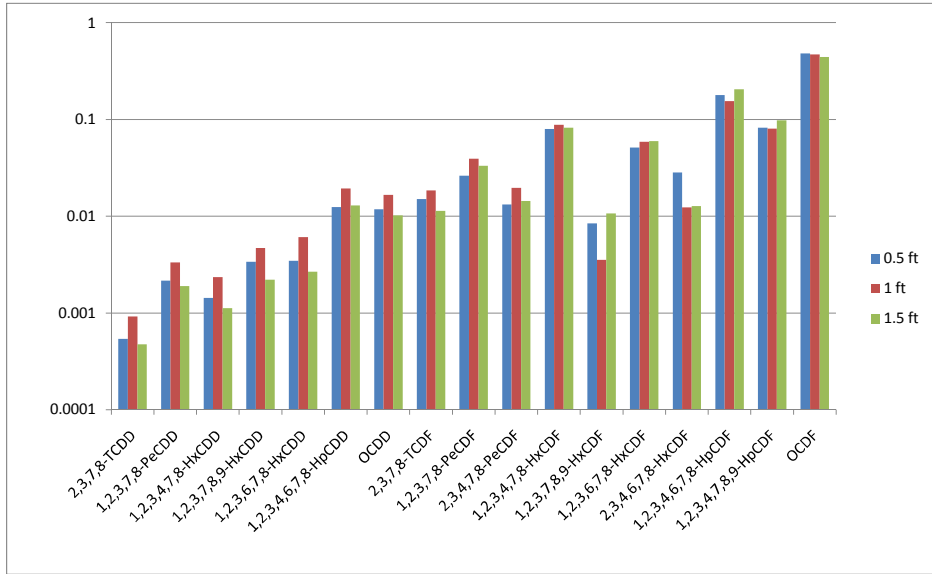
Set 1 - Dioxin/Furan profiles for select soil samples

Area 4
 Grid Q-5
 Sample SA156-0.5B
 TTEQ (ppt) 1,300

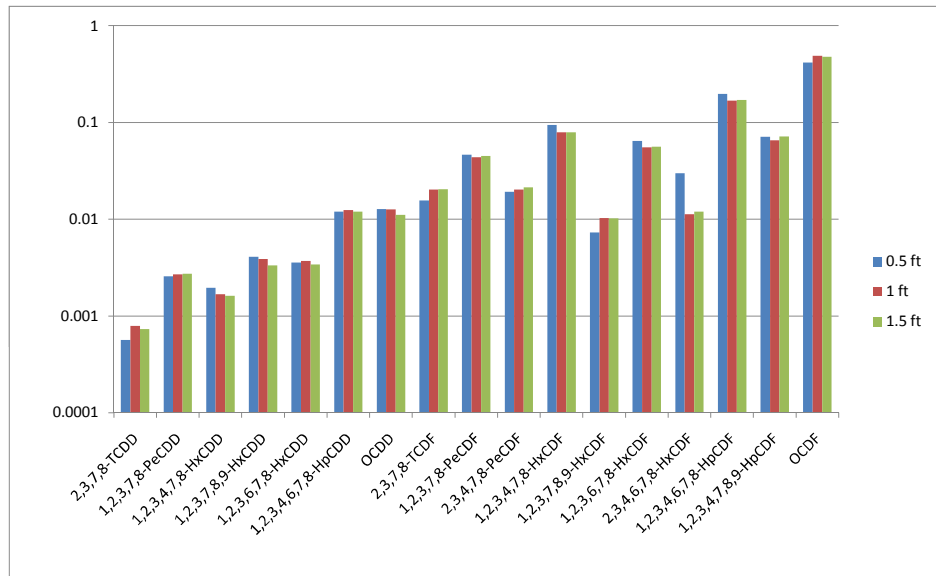


Set 2 - Dioxin/Furan Congener Profiles for select supplemental (depth) soil samples

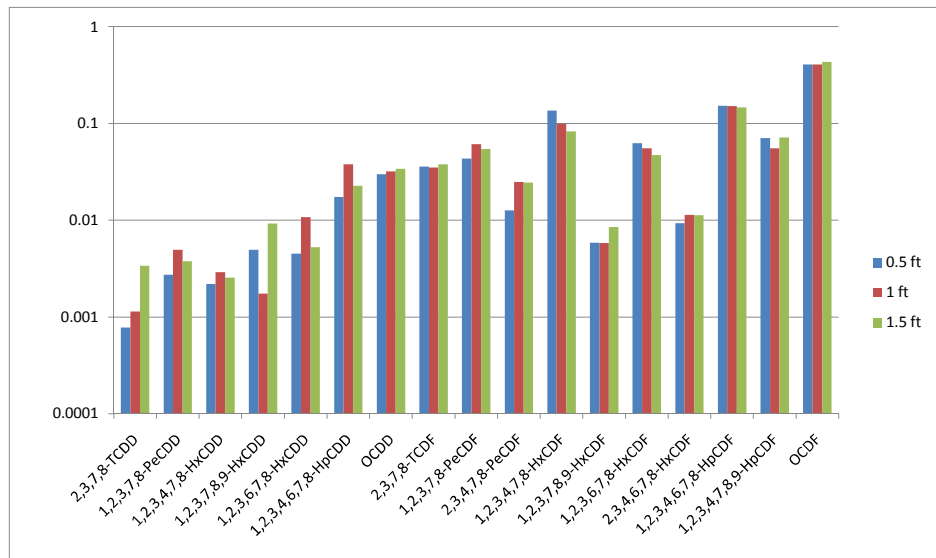
Location SA201-0.5B
 Area 1 - Near Berm
 Grid I-3



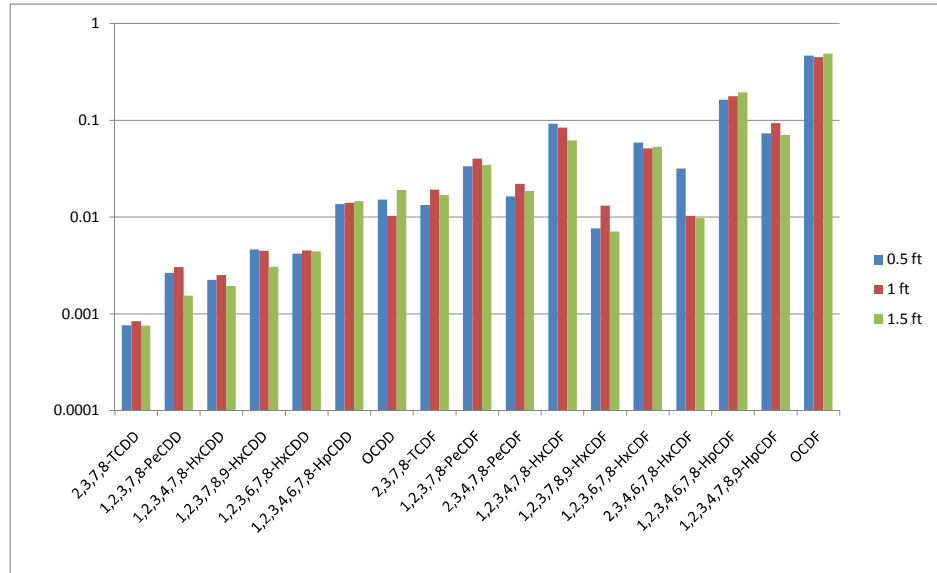
Location RSAH3-0.5B
 Area 1
 Grid H-3



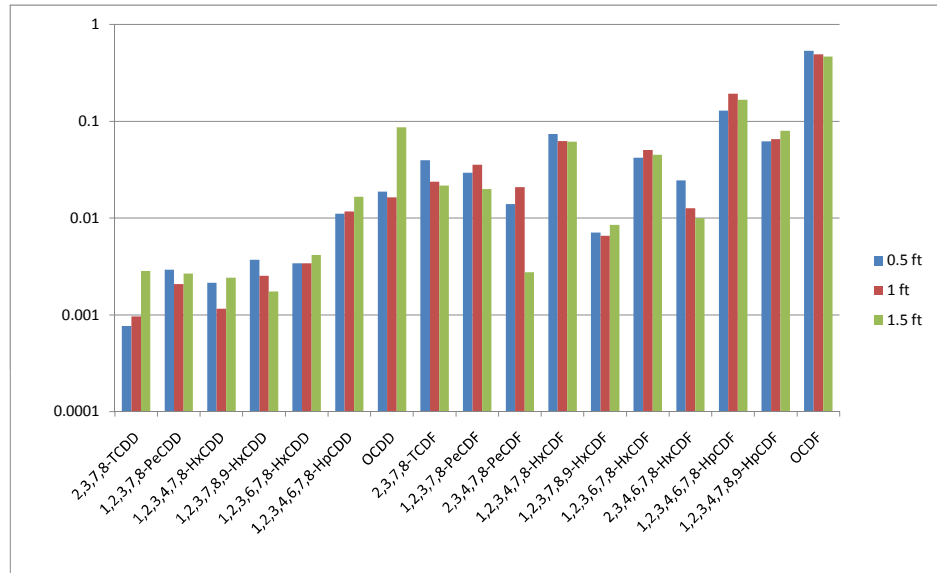
Location RSAL3-0.5B
 Area 1
 Grid L-3



Location SA155-0.5B
 Area 2 - Ditch
 Grid M-7



Location SA104-0.5B
 Area 2
 Grid M-5



Location SA200-0.5B
 Area 2
 Grid O-6

