APPENDIX A

Sample Collection, Laboratory, and Quality Assurance/Quality Control (QA/QC) Procedures

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A. Sample Collection Procedures

1. Soil Sampling

ENVIRON attempted to locate individual soil sampling borings at the approximate geometric center of each pond cell and the centerlines of the ditches. The coordinates of each sample location was logged in the field using hand-held Global Positioning System equipment (GPS). Surficial and subsurface samples were collected with a direct-push drill rig (i.e., Geoprobe) using wide-bore (2-inch or greater) sample spoons. Soil cores were visually logged in accordance with ASTM D-2488 (ASTM 1990) by an experienced ENVIRON geologist or engineer.

All subsurface tools used in the collection of soil samples were constructed of stainless steel. All sampling equipment was decontaminated upon arrival at the site, between all sampling locations, at the end of each day of sampling operations, and prior to demobilization from the site. Equipment decontamination procedures include the following: thoroughly cleaning using a high pressure steam cleaner and/or rinse with tap water; rinsing and scrubbing with non-phosphate detergent solution, rinsing with tap water to remove detergent, and rinsing with distilled/deionized water.

2. Ground Water Sampling

Ground water samples were collected from existing monitoring wells using either down-hole pumps with dedicated tubing or disposable, dedicated Teflon bailers, because of low-flow conditions (at certain locations the pumping caused the wells to become dry). The type of sampling method used at each ground water monitoring location is provided in the well logs (Appendix B). Monitoring wells were purged of three well volumes (plus filter pack volume) or until field parameters (temperature, specific conductivity, dissolved oxygen, and oxidation-reduction potential, and pH) stabilized to three consecutive readings within 10 percent of the arithmetic mean of the values. No permanent ground water monitoring wells were constructed as part of the characterization. Similar to the procedures applied for soil sampling equipment, all equipment used in the collection of ground water samples was decontaminated upon arrival at the site, between all sampling locations, at the end of each day of sampling operations, and prior to demobilization from the site. Equipment decontamination procedures included the following: thoroughly cleaning using a high pressure steam cleaner and/or rinse with tap water; rinsing and scrubbing with non-phosphate detergent solution, rinsing with tap water to remove detergent, and rinsing with distilled/deionized water.

B. Field QA/QC Procedures

QA/QC sampling procedures were employed during the field program to ensure the reproducibility of results, determine the effects of sample shipping and handling, and assess the quality of field decontamination procedures. The QA/QC procedures applied in the field are discussed in the following sections.

1. Field Duplicate

Field duplicates were collected and submitted to the laboratory at a rate of one duplicate per 10 soil borings or monitoring wells, with a minimum of two per medium. Duplicate samples are used to assess precision; therefore, both pairs of the duplicate samples were collected in the same location, in the same exact manner, and at the same time. The specific locations of field duplicates included P-17 in the Northern Exposure Area, and P-5 and P-7 in the Southern Exposure Area.

2. Rinseate (Field) Blanks

One rinseate blank was collected for every 10 soil borings and monitoring wells, with a minimum of at least two equipment rinseate blanks collected in the field. These samples are used to test for residual contamination of the sampling equipment. The rinseate blanks were be collected by pouring de-ionized water over decontaminated sampling equipment into labeled sample bottles. Pre-cleaned sample bottles with appropriate preservatives were supplied by the analytical laboratory. Immediately after filling, the sample bottles were securely closed, placed in a cooler, and kept chilled until delivery to the analytical laboratory. Equipment rinseate blanks were considered associated with all samples collected since the last equipment rinseate blank was collected. Three rinseate blanks were collected in the field (identified as WB01, WB02, and Rinse 3); however, the sample container for WB02 broke in transit to the laboratory.

3. Trip Blanks

One trip blank was collected per cooler used to ship VOC samples. Trip blanks were prepared and shipped by the analytical laboratory with the sample containers. These samples are used to identify contamination introduced in the field or by the laboratory. Trip blanks were labeled in the field at a specific sample location, and shipped to the analytical laboratory with the associated samples. One trip blank sample was included in

A-2

each shipment to the analytical laboratory. Trip blanks are considered to be associated with all samples in the same shipment to the laboratory.

C. Sample Custody and Shipping Procedures

1. Field Custody Procedures

Field personnel are responsible for recording field activities on the appropriate field documentation form in sufficient detail to allow the event to be reconstructed without relying on memory. It is the responsibility of the Field Activities Manager to ensure that all documents are complete and legible. At the end of each day, all documents completed that day were reviewed by the Field Activities Manager for accuracy, completeness, and legibility.

The following field documentation forms or records were used during the field activities, at a minimum:

- Sample Chain-of-Custody record;
- Soil Sampling Log;
- Field Activity Daily Log;
- Sample Alteration Form; and
- Corrective Action Report.

Each completed form (a copy or original depending on the type of form) was maintained on the site in chronological order with other completed forms of the same type until the completion of the field activity. Copies of the chain-of-custody forms are provided at the end of this appendix.

2. Field Activity Daily Log

A field activity daily log was used as a record of daily field activities showing the sequence of events. The log included the following information:

- Project name and number;
- Date;
- Starting/ending time and nature of each major field activity;
- Names of all personnel on the site (including visitors), and a description of their involvement with the tasks being conducted;
- Weather conditions;
- References to appropriate field logs for details of each activity performed (e.g., reference sample collection logs for details of all samples collected that day);

- Identification of any photographs taken; and
- Signature of Task Leader or other reviewer.

The Field Activities Manager was responsible for ensuring that all activities were documented in the field activity daily log and that the details of each activity are recorded on the appropriate field documentation form.

3. Photographic Documentation

Color photographs were taken of representative sampling locations and the surrounding site to show the area, sampling equipment, and related site activities. ENVIRON has retained these photographs on file, but is not including them as part of this report.

4. Sample Identification and Labeling Procedures

Each sample collected for testing was assigned a unique sample identification (ID) code. The sample ID facilitates data management by referencing the analytical laboratory, depth interval, site, date, and location. All samples were labeled with the sample ID code and other field information. Duplicate and QA/QC samples (i.e., field equipment rinseate blanks and trip blanks) were similarly labeled.

The sample labels were placed on the sample containers so as not to obscure any data on the containers. Sample information was printed on the labels in a legible manner using waterproof ink. The label contained sufficient information so the sample could be identified on the sampling information form or sample collection log. Sample labels contained the following information:

- The project name and number
- A unique sample identification.
- The date and time.
- Identification of preservatives used, if any.
- A list of analytical tests to be performed on the sample.
- Other necessary remarks.
- Name of the sampler.

5. Chain-of-custody Record

The chain-of-custody record for each sample originates at the site, beginning with sample collection, and is completed prior to shipment to the laboratory. A copy of the chain-of-custody record accompanies the sample to the laboratory in order to establish the

documentation necessary to trace sample possession from sample collection through sample analysis. The sampling portion of the chain-of-custody record contained:

- List of sampling team members;
- Sample number;
- Signature of sampler or bottle preparer;
- Date and time of sample collection;
- Sample depth;
- Medium type;
- Signatures of persons involved in the chain of possession;
- Inclusive dates of possession; and
- Preservation.

6. Shipping Procedures

The following procedures were followed for packing samples for shipment to the laboratory:

- All sample container caps were checked for tightness.
- The sample containers were placed in coolers, allowing sufficient space for the addition of packing material between the sample containers.
- Ice packs (or equivalent) were placed on top of and between the samples.
- A copy of the chain-of-custody form was placed in a sealed, clear plastic envelope and placed in the cooler.
- Custody seals were placed on the outside of each cooler.
- The shipping coolers were taped shut.

Samples were shipped every one or two days to the laboratory via overnight courier.

D. Laboratory Custody Procedures

The laboratory Group Leader accepted custody of the samples shipped from the field and verified that the information on the sample label matches the information on the chain-of-custody record. Pertinent information relating to shipment, pickup, and courier were also be verified on the chain-of-custody record.

The Sample Receipt/Sample Entry Group Leader entered the appropriate data from the chain-of-custody record into the laboratory sample tracking system (both a written file and an electronic database) using the sample number from the sample label or assigning a unique laboratory number to each sample. The Sample Support Group Leader transferred the samples to

the proper analyst, stored the samples in the appropriate secure area, and documented in writing all internal transfers of the samples.

The Sample Receipt/Sample Entry Group Leader notified ENVIRON's Field Activities Manager of any discrepancies noted on the chain-of-custody or sample labels. Samples were not analyzed until the discrepancy was resolved. Any changes made were documented by the laboratory and ENVIRON personnel.

The Sample Receipt/Sample Entry Group Leader and the Sample Support Group Leader are responsible for custody of samples from the time they are received until sample analysis is completed. Any unused portions of samples remaining after completion of analysis by the laboratory was disposed of in accordance with procedures developed by the laboratory and consistent with applicable laws and regulations governing sample disposal.

The laboratory portion of the chain-of-custody form was completed by the designated laboratory sample custodian and contains:

- Name of person receiving the samples;
- Laboratory sample number;
- Date of sample receipt by the laboratory;
- Analyses requested; and
- Sample condition and temperature.

Immediately upon arrival at the contract laboratory, the laboratory recorded the condition of the shipping container and sample containers. The original chain-of-custody form was returned from the laboratory as part of the final analytical report to ENVIRON.

E. Laboratory QA/QC Procedures

In addition to performing the analysis of samples, the analytical laboratory is responsible for performing several quality assurance/quality control (QA/QC) procedures. Laboratory QC checks are accomplished through the use of system checks and QA/QC samples that are introduced into the sample analyses stream. Laboratory system checks and QA/QC samples are required by the selected USEPA analytical methods. Laboratory QA/QC checks were performed and samples were analyzed at the frequencies stated below or at the frequencies established by appropriate USEPA analytical methods, whichever is greater. The QC check samples are listed and defined below.

1. Laboratory Calibration Procedures

The laboratory calibrates its analytical instruments by establishing analytical curve based on the absorbance, emission intensity, or other measured characteristics of known standards. The calibration standards must be prepared using the same type of acid and at an equivalent concentration as used in the sample preparation. Initial instrument calibration should consist of analysis of analytical standards for a series of different specified concentrations, used to define the quantitative response, linearity, and dynamic range of the instrument to target compounds. Continuing calibration should consist of an analytical standard run every twenty analytical samples or every twelve hours, whichever is more frequent, to verify the calibration of the analytical system.

A calibration blank is prepared by the laboratory using acidified distilled/deionized water to ensure that contamination is not present in the preparation water or in the analytical instrument due to carry over from other samples or standards. The initial calibration blank (ICB) is analyzed after the analytical standards, but not before analysis of the initial calibration verification (ICV) solution(s), during the initial calibration of the instrument. A continuing calibration blank (CCB) is analyzed after every initial and continuing calibration verification. The CCB shall be analyzed at a frequency of 10% or every twelve hours during the run, whichever is more frequent.

2. Method Blank

The method blank is used to detect any contamination introduced by the laboratory. A method blank is a quality control sample prepared by the laboratory that contains distilled/deionized water and the same reagents used with the field samples and carried through the entire analytical procedures (digested and analyzed). An aqueous method blank is treated with the same reagents as a sample with a water matrix; a solid method blank is treated with the same reagents as a soil sample. Method blanks were generated and analyzed at a frequency of at least one per twenty samples of a given matrix (e.g., soil or water).

3. Matrix Spike/Matrix Spike Duplicates

The laboratory prepares a matrix spike (MS) sample by introducing a known amount of chemical to the matrix and subjects the sample to the same analytical procedures as field samples of the matrix. The process is repeated for a matrix spike duplicate (MSD). From this analysis, the laboratory determines the percent recovery (PR) and the relative percent difference (RPD) in recovery between the MS and MSD. The laboratory runs MS and MSD to determine long-term precision and accuracy of an analytical method on various matrices and to demonstrate acceptable compound recovery by the laboratory at the time of sampling.

4. Laboratory Control Sample

The laboratory analyzed laboratory control samples (LCS) periodically during the analysis of field samples to assess the accuracy of the analytical method and the laboratory's performance. In addition, the laboratory analyzes an LCS duplicate and calculates PR for the LCS and LCS duplicate and the RPD for the two samples. The PR and RPD for the LCS are reported by the laboratory as part of the analytical data package. The laboratory compares the PR and RPD to acceptable ranges for these values in the laboratory's SOP.

5. Surrogates

Surrogate compounds, which are chemicals that are not expected to be detected in field samples, are added to certain samples (VOCs, semivolatiles, PCBs, and pesticides) in known quantities to determine recovery for the purpose of determining analytical efficiency. The PR of the surrogate is calculated by the laboratory and compared to the accepted range of PRs in the laboratory's SOP.

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DISTRIBUTION: WHITE - Stays with the Sample; CANARY - Returned to Client with Report; PINK - Field Copy

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DISTRIBUTION: WHITE - Stays with the Sample; CANARY - Returned to Client with Report; PINK - Field Copy

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DISTRIBUTION: WHITE - Stays with the Sample; CANARY - Returned to Client with Report; PINK - Field Copy

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DISTRIBUTION: WHITE - Stays with the Sample; CANARY - Returned to Client with Report; PINK - Field Copy

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Severn Trent Laboratories, Inc.

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DISTRIBUTION: WHITE - Stays with the Sample; CANARY - Returned to Client with Report; PINK - Field Copy

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